



Effect of NaCl and pH on Bioelectricity Production from Vegetable Waste Extract Supplemented with Cane Molasses in Dual Chamber Microbial Fuel Cell

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ABSTRACT

Vegetable waste has been analyzed for bioelectricity production in locally designed U-shaped microbial fuel cell. These fuel cells were subjected to different concentrations of NaCl and pH for evaluation of power generation. The MFC with 50 mM NaCl concentration showed maximum response and gave 88.9 W/m² power densities and 569 mV of open circuit voltage. The results showed that NaCl, at its optimum level, plays an important role in increasing the bioelectricity and sustaining the internal resistivity. Six different pH parameters were subjected to check the effectiveness of the treatment on the power production. The maximum power production achieved at pH 4.5 i.e. 70.5 W/m² of power density at external load of 200 Ω . Which reveals that if the pH difference between the two chambers is increased the power density increases. Bioelectricity production from molasses has not been studied considerably. Present study describes the use of molasses with different organic loads in MFC-1, MFC-2 and MFC-3 with 10, 15 and 20% of sugar contents, respectively, for bioelectricity production. These MFCs were designed locally and maximum power densities of 72820mW/m², 104400mW/m² and 44340mW/m² for MFC-1, MFC-2 and MFC-3, respectively were obtained at 400 Ω external load. The columbic efficiencies were 73.9%, 81.7% and 63.2%, respectively.

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Authors' Contribution

MMJ supervised the experimental work. MAN and MUA performed the experimental work. MMJ and MUA wrote the manuscript.

Key words

Vegetable extract, Microbial fuel cell (MFC), Bioelectricity, Cation exchange membrane, Acidophilic and alkaliphilic

INTRODUCTION

Microbial fuel cells (MFCs) are the alternative source of energy which is gaining attention day by day (Logan *et al.*, 2006). These are bio-electrochemical devices which uses microbial culture for the electricity production. The microbial flora generates electrons, by utilizing various organic substrates, which is used for the bioelectricity production as explained by (Oh *et al.*, 2004). The substrates that have been reported to use as organic source in the microbial fuel cell are of many types. These substrates could be simple compounds like sucrose, glucose, fructose ethanol and acetate (Logan *et al.*, 2007; Jafary *et al.*, 2013) or it may also be complex media like food waste (Goud *et al.*, 2011; Li *et al.*, 2013a) and different waste water including domestic, agriculture industrial waste water etc. (Nimje *et al.*, 2012).

Ionic strength is one of the important aspect due to

which the internal resistance (R_{int}) in the microbial fuel cell is affected. Early studies done by Li *et al.* (2013b) showed that by increasing the internal resistance the power densities are decreased. In some other studies it has been suggested that increasing the NaCl concentration decrease the internal resistivity by improving the ionic strength and increase the energy production as explained by Kyung *et al.* (2004) and Lefebvre *et al.* (2012). Some studies conducted by Huang *et al.* (2008) also suggested that as the micro flora in the anodic chamber varies the salt tolerance may affect the bioelectricity production.

The bioelectricity production through MFC is greatly effect by the pH of the anodic chamber (He *et al.*, 2008). It has been found, by changing the pH of the substrate in anodic chamber, that many physiological changes may occur like ionic strength, membrane potential, microbial metabolism and proton generation potential (Zhang *et al.*, 2010; Jung *et al.*, 2011). The ionic strength of the MFC can be improve by adjusting the pH of the anode chamber which can increase the power output by decreasing the internal resistance (Li *et al.*, 2013b). In case of the dual chamber microbial fuel cell it has been found the higher pH difference in the two chambers can increase the power

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density (Jadhav and Ghangrekar, 2009). If alkaliphilic bacteria have been used the high substrate pH and low cathode pH has been found to give the maximum power output (Liu *et al.*, 2010; Zhuang *et al.*, 2010) while in some other study it has been showed that lower substrate pH in anode give maximum potential (Rikame *et al.*, 2012).

The substrate used in the MFC is one of the important factors which need to be optimized for higher power output. Food waste was also in interest of many researchers which serves as rich carbon and energy source for microorganism and also consists of almost 30% of the municipal waste. Huge variety of substrates have been used in the MFC as carbon sources for microbial culture such as glucose and fructose (Jafary *et al.*, 2013), waste water (Nimje *et al.*, 2012; Akman *et al.*, 2013) and food waste (Goud *et al.*, 2011; Li *et al.*, 2013a). Molasses on the other hand not been extensively studied but some reports from past years showed molasses an effective electron donor in microbial fuel cells (Niessen *et al.*, 2004; Fernando *et al.*, 2014; Sevda *et al.*, 2014).

MATERIALS AND METHODS

Substrate preparation

Vegetable waste from house hold kitchen was collected which included spinach leaves, cucumber and potato leaves, carrot and cabbage. These were converted into small pieces and blended with phosphate buffer solution (0.1 M) with pH 4.5 in 1:1 to solid weights of the waste pieces. The slurry was taken and filtered for further use.

Enrichment of microbial culture for inoculum preparation

Sewage sample was taken from the sewage waste water from Haal road, Lahore. Designed synthetic waste water (DSWW) was prepared as demonstrated by Venkata Mohan *et al.* (2010). The composition of DSWW is: K_2HPO_4 -0.25g/l, KH_2PO_4 -0.25g/l, NH_4Cl -0.5g/l, $ZnCl_2$ -11.5mg/l, $MgCl_2$ -3g/l, $CoCl_2$ -25mg/l, $CaCl_2$ -5mg/l, $CuCl_2$ -10.5mg/l, $MnCl_2$ -15 mg/l, $NiSO_4$ -16mg/l, $FeCl_3$ -25mg/l, glucose-3g/l and Vitamins-5g/l. The DSSW media was purged with nitrogen to provide anaerobic environment. The sample was inoculated in the DSWW media and incubated for 72 hours at 37°C. The enriched culture was centrifuged (6000 rpm at 20°C) and the pellet was washed thrice with saline phosphate buffer solution with 0.85% NaCl.

Microbial fuel cell configuration

Six U-shaped dual chambered MFCs with cation exchange membrane (CMI 7000) were prepared as explained in our previous study. All MFCs were

supplemented with 200 ml substrate in anode but each with different NaCl concentration i.e. MFC-1 (25 mM), MFC-2 (50 mM), MFC-3 (75 mM), MFC-4 (100 mM), MFC-5 (125 mM) and MFC-6 (150 mM). Tap water was used as catholyte which already contains enough salts and traces for the process to run. For optimization of pH studies, six different MFCs were set at pH: 3.5, 4.5, 5.5, 6, 7 and 7.5. NaCl concentration was kept 50mM in all MFCs. Graphite rods were used as anode and cathode, three in each chamber, with total surface area of $3 \times 0.0015 \text{ m}^2$. The electrodes were connected to a multimeter (UNI-T UC- 60A), with personal computer (PC) interface, parallel to variable resistance (5-5000 Ω) as shown in Figure 1.

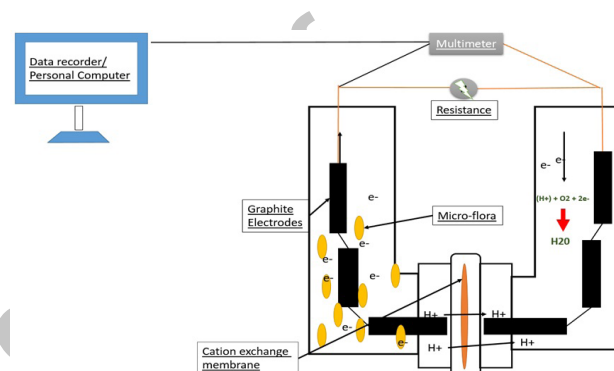


Fig. 1. The diagrammatic presentation of the U-shaped MFC designed for this project.

Experimental design for utilization of molasses as substrate for bioelectricity production

Clarification of cane-molasses was performed by using the method of Panda *et al.* (1984). MFC-1 (10% sugar content with 9119mg/l COD), MFC-2 (15% sugar with 12076mg/l COD) and MFC-3 (20% sugar content with 15390mg/l COD) were set up for this experiment. NaCl (50mM) was added to provide sufficient ionic strength to media in all three MFCs. Cathode of MFC was supplemented with tap water (pH 7.0) whereas clarified molasses (pH 4.5) was added in anode chamber of MFC, along with mix microbial culture enriched in synthetic waste water media as inoculum (Mohan *et al.*, 2010).

Data analysis

Data was recorded across different resistance from 5-5000 Ω and the current was calculated by relation V/R where V is voltage and R is respective resistance and power was calculated using the formula $P=VI$, where I represents current. The power density and current density were calculated by dividing the electrode (anode) surface area to power and current, respectively. The slope of the linear zone of the graph between the current and voltage

gives the internal resistance (Logan *et al.*, 2006). Columbic Efficiency (C.E) was also calculated by checking the COD difference after specific time period. Following relation was used for the C.E in which ΔCOD is change in chemical oxygen demand, I is current, V is substrate volume in anode, b is total number of electrons i.e. 4 per mol of oxygen, F is faraday's constant, and M is molar weight of oxygen.

$$CE = \frac{M \int_0^{tb} I dt}{FbV\Delta COD}$$

RESULTS AND DISCUSSION

Performance of MFC at different NaCl concentration

The result from two ranges of the salt treatments, i.e. lower range (25 to 75 mM) and higher range i.e. (100 to 150 mM), showed that with higher salt concentration the microbial fuel cell performance decreased which is the evidence of lower microbial activity and increased internal resistivity (Fig. 2) due to volumetric resistance as described by Clauwaert *et al.* (2008). Li *et al.* (2013b) also suggested that NaCl concentration improves the ionic strength which helps in electron and proton transfer through the microbial fuel cell. The internal resistance of the higher range treatments is steady but with slight changes with time as compare to lower treatments which showed to be the reason of higher microbial activity in the lower range NaCl concentration and vice versa which could be the optimal requirement for microorganism as explained by Prescott (2002). The higher microbial activity also leads the MFC to increase the power density which is why higher power output was noted at lower range of salt concentration (Fig. 2). The results confirms the optimum concentration of salt should be needed for efficient production of bioelectricity and MFC performance which was also previously studied by Li *et al.* (2013a) which might be different for a MFC depending on the substrate and microbial culture used.

The current was calculated by observing the voltage on the external load as described by Logan *et al.* (2006). It was found that 200 Ω of external resistance provides maximum power density which was to be selected for analysis. The maximum power production was 88.99 W/m² at 50 mM NaCl concentration on the second day of the process while 67.78 W/m² and 81.12 W/m² power densities was for the first and third day, respectively (Fig. 2). The power output from this work is higher, to our knowledge, than most of the reports by Wang *et al.* (2014); Yong *et al.* (2014); Cao *et al.* (2015). The concentration of 50 mM then other concentration confirms the optimized condition for running the MFC for this work. Although other lower ranged concentration was also quite significant and closer

each other in term of the open circuit voltage and power density (Figs. 1 and 2). The lower range support higher power output because of efficient microbial activity and lower internal resistance as compare to the higher range of the NaCl concentration as described by Li *et al.* (2013b).

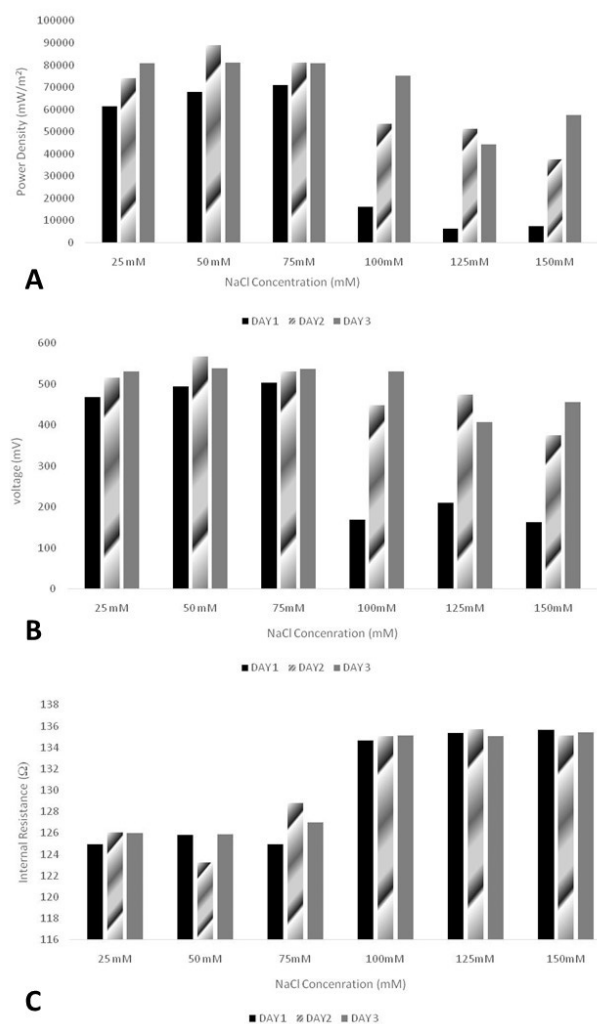


Fig. 2. Effect of concentration of NaCl on maximum power density obtained at 200 Ω (A), Open circuit Voltage for 3 days (B) and internal resistivity (C).

Effect of pH on power generation in MFC

By keeping the cathodic chamber pH constant, i.e. 7 to 7.5, the anodic chamber pH is being decreased in which the increase in the power generation occur gradual decrease in the pH up to a certain limit. Changes in the pH may result various physiological changes in the Microbial fuel cell like changes in membrane potential, microbial metabolism and ionic strength especially (Zhang *et al.*, 2010; Jung *et al.*, 2011). The increase in power with

decrease in pH may due to the creation of pH gradient between the anodic and cathodic chamber which resulted the flow of ionic movement from one to another chamber efficiently (Jadhav and Ghangrekar, 2009; Zhuang *et al.*, 2010). Although the result was in contradiction with the reports of Zhuang *et al.* (2010) whose power output was higher at higher anodic and lower cathodic pH but in this case higher potential was at pH 4.5 with power density of 70.5 W/m^2 (Fig. 3) which was higher than reported by Zhuang *et al.* (2010) but when the pH is decreased to 3.5 the power and voltage generation was getting low (Fig. 3) which indicated the decrease in the microbial activity with respect to the conditions (Gaboriaud *et al.*, 2006). In some other reports the acidic pH of anode chamber gave higher power output (Rikame *et al.*, 2012; Li *et al.*, 2013b).

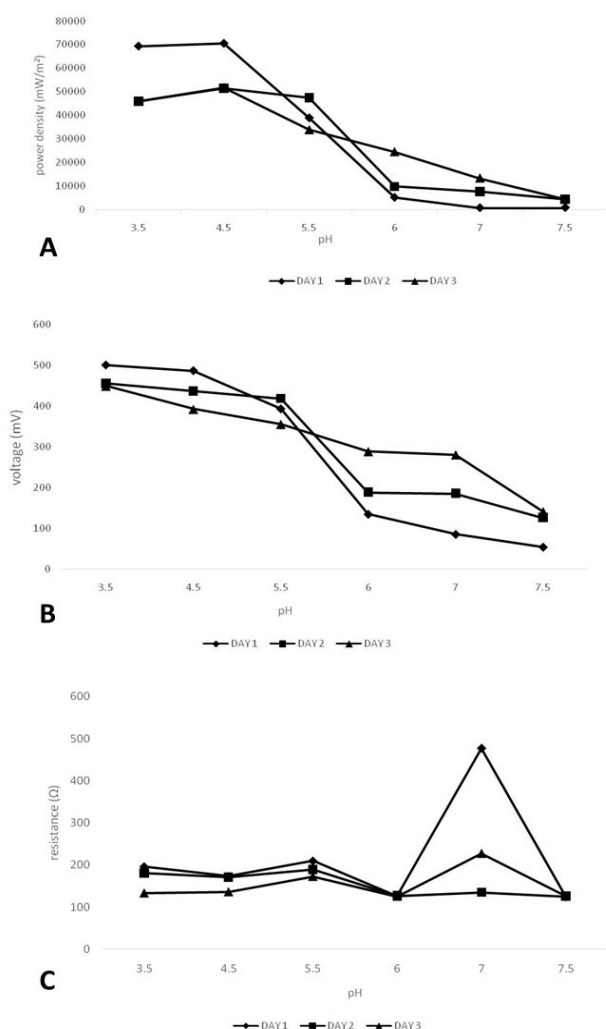


Fig. 3. Effect of different pH on maximum power density (A), open circuit voltage recorded for 3 days (B), and internal resistivity of the MFCs (C).

The MFCs with lower range pH i.e. MFC-1, 2 and 3 with pH 3.5, 4.5 and 5.5, respectively gave maximum power output and open circuit voltage (Fig. 3) due to higher pH difference between cathodic and anodic chamber (Zhang *et al.*, 2010; Rikame *et al.*, 2012; Li *et al.*, 2013b). The higher power density was 70.5 W/m^2 observed at the first day of the process in the MFC-2 with pH 4.5 (Fig. 3). While the second and third day was also higher than the rest of the MFCs which was almost 51 W/m^2 except to the first day process of MFC-1 which was 69.3 W/m^2 . The result was higher than most of the other reports to our knowledge (Li *et al.*, 2013b; Wang *et al.*, 2014; Yong *et al.*, 2014; Cao *et al.*, 2015). As the pH gets closer to the cathodic pH which was 7 to 7.5 the lower power output was observed. It also has been observed that at pH 6 and 7.5 the internal resistance was minimum (Fig. 3) which is assumed to be the reason of optimized condition for ionic strength for minimum internal resistance with respect to the cathodic chamber (Li *et al.*, 2013b). The results showed that MFCs with lower ranged pH gave higher potential but maximum ohmic losses as compare to those with higher pH treatments with one unit difference as referred to cathode pH (Clauwaert *et al.*, 2008).

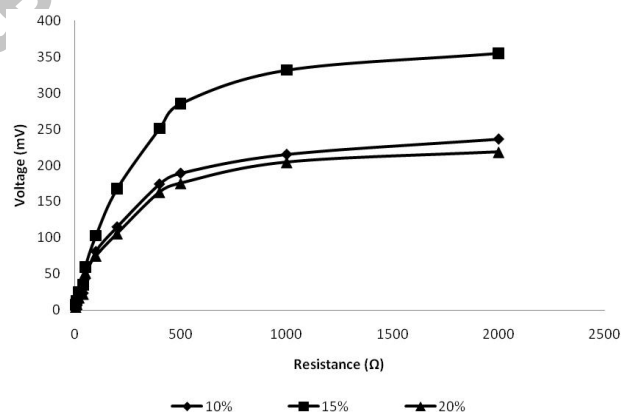


Fig. 4. Graph between voltage and resistance.

Efficiency of microbial fuel cell and electricity generation with molasses as substrate

It has been found that the C.E of MFC-2 (81.7%) was higher, compared to MFC-1 (73.9%) and MFC-3 (63.2%). Lower efficiency of the MFC-1 and MFC-3 can be reasoned due to lower microbial metabolic rate, for which higher internal resistance comparatively to the current production might reduced the coulombic efficiency. Same pattern of results has been reported by Rikame *et al.* (2012) and Juang *et al.* (2011).

Voltage was analyzed on variable external load and a straight line was observed up to 400Ω for MFC-1 and

MFC-3 while in case of MFC-2, straight line goes to 500 Ω , which shows that higher power density can be obtained at these external loads (Fig. 4). MFC-1 recorded maximum power density (72820mW/m², at 400 Ω) as compared to MFC-3 (44340mW/m², at 500 Ω) and MFC-2 (104400mW/m², at 500 Ω) (Fig. 5). Internal resistance of MFC-2 was higher as compare to other two i.e. 283.3 Ω (Fig. 6) but higher power and electric-current output is the evidence of the high activity of microorganisms comparatively. The results of power density in present study was higher than most of the reports using the molasses as electron donor (Niessen *et al.*, 2004; Fernando *et al.*, 2014; Sevda *et al.*, 2014).

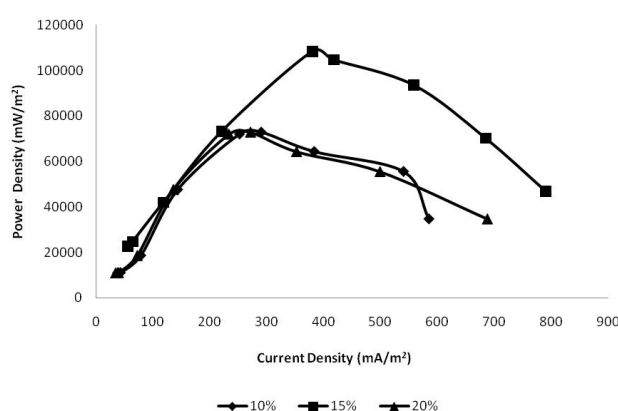


Fig. 5. Polarization curve of power density and current density

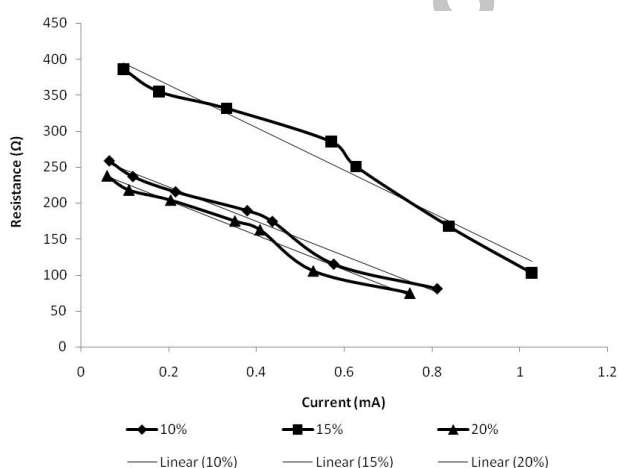


Fig. 6. Internal resistivity from resistance and current slope of the MFCs.

Kargi and Eker (2007) used molasses with urea and reported power density of 2.6mW/m². While in comparison to other substrates, molasses was found to be an effective

substrate in term of power production. Mansoorian *et al.* (2013) reported power density of 230mW/m² from food processing waste water. Whereas, relatively higher power densities of 590mW/m² (xylose as substrate) and 1500mW/m² (chocolate industry wastewater as substrate) has been reported by Mäkinen *et al.* (2013) and Patil *et al.* (2009), respectively.

Results of present work also favor open circuit voltage of MFC-2 with average value of 471mV while MFC-1 and MFC-3 had 338mV and 312mV of potential difference, respectively (Fig. 7). Open circuit voltage recorded in present work is lower than some reports in which different kinds of substrate have been used (Zhuang *et al.*, 2010; Rikame *et al.*, 2012; Wang *et al.*, 2014). This could be possibly explained by the existence of higher volumetric resistivity in cane molasses as compared to other substrates (Clauwaert *et al.*, 2008).

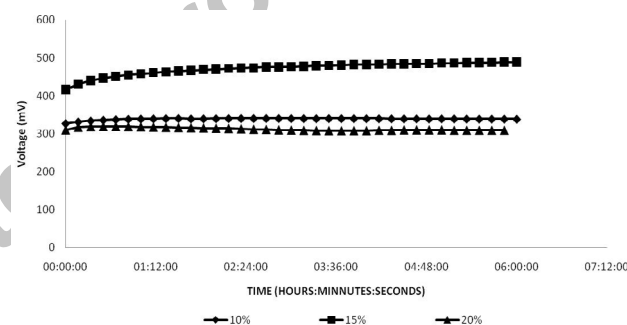


Fig. 7. Trend of open circuit voltage with time period.

CONCLUSION

Sodium chloride is common salt that is been used in homes gave us most economic choice to use as ionic agent for increasing the ionic strength in the Microbial Fuel Cell. The optimum concentration of the salt can increase the power density by minimizing the internal resistance. Increasing the salt concentration from the optimum level might result in lower power output due to increased internal resistance and lower microbial metabolic rate to produce electrons. It has been found that pH is a crucial element in the optimizing and increasing the power of the microbial fuel cell. If the microbial fuel cell is dual chamber it should be necessary to increase the pH difference up to a limit between the two chambers for ionic facilitation and higher voltage production. The choice of microorganism should be more like acidophilic for low anode pH or alkaphilic for high anode pH, to support other findings.

Present study also revealed that molasses can act as good initiating agent or carbon source for microorganism in microbial fuel cells for bioelectricity production.

Literature reports suggested the use of other substrates for bioelectricity production but problem of lower internal resistivity with these substrates empowers the fact of molasses as potent substrate in MFCs.

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Statement of conflict of interest

The authors have declared no conflict of interest.

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Uncorrected Proof