

Review Article

The Point behind Translation of Aptamers for Point of Care Diagnostics

Tarun Kumar Sharma^{1,2*}, John G. Bruno³ and William C. Cho⁴

¹Centre for Biodesign and Diagnostics, Translational Health Science and Technology Institute, Faridabad, Haryana-121001, India; ²AptaBharat Innovation Private Limited, Translational Health Science and Technology Institute Incubator Haryana-121001, India; ³Operational Technologies Corporation, 4100 NW Loop 410, Ste, 230, San Antonio, TX 78264, USA; ⁴Department of Clinical Oncology, Queen Elizabeth Hospital, Hong Kong.

Abstract | Owing to the high affinity and specificity in recent years aptamers, have emerged as competitive tools with the potential to replace antibodies in many diagnostic formats. The aptamer industry is continuously growing to create space in research and diagnostics markets. However, translation of aptamer in several applied disciplines is still lagging behind as compared to antibodies. In this article, we discuss the translational aspects of aptamers, current potential of aptamers and strategies to create the “bench-to-bedside” translation of aptamer technology.

Editor | Dr. Muhammad Munir, The Pirbright Institute, Woking, Surrey, UK.

Received | September 01, 2016; **Accepted** | October 17, 2016; **Published** | November 02, 2016

***Correspondence** | Tarun Kumar Sharma, Centre for Biodesign and Diagnostics, Translational Health Science and Technology Institute, Faridabad, Haryana-121001, India; AptaBharat Innovation Private Limited, Translational Health Science and Technology Institute Incubator Haryana-121001, India. **Email:** tarun@thsti.res.in, aptabharat@gmail.com.

Citation | Sharma, T.K., J.G. Bruno and W.C. Cho. 2016. The point behind translation of aptamers for point of care diagnostics. *Aptamers and Synthetic Antibodies*, 2(2): 36-42.

Keywords | Aptamers, Translation of aptamer, Aptamer industry, Point of care diagnostics, SELEX

Aptamers are single stranded DNA (ssDNA) or RNA molecules that are generated through a process of *in vitro* evolution called Systematic Evolution of Ligands by EXponential Enrichment (SELEX) (Tuerk and Gold, 1990). By virtue of their complex 2D and 3D structures, an aptamer binds to its cognate target with high specificity and affinity (Blind and Blank, 2015; Lange et al., 2012; Sharma et al., 2014; Weerathunge et al., 2014). These receptor-like molecules have emerged as strong rivals of antibodies, which are long-standing tools used to capture antigens (Chopra et al., 2014; Sharma and Shukla, 2014a, b). Development of a rapid antigen detection assays always requires specific capture of the ligand. Anti-ligand antibodies are commonly used to specifically capture ligands from various clinical samples including, but not limited to blood, serum, cerebrospinal fluid (CSF) etc. (Citartan et al., 2012; Gopinath et al., 2016). However, antibodies have to be raised in

animals, are prone to batch-to-batch variation due to differences in titers elicited in different hosts, and the antibody binding kinetics (K_{on} and K_{off}) or on and off-rates cannot be controlled (Bruno, 2014; Bruno and Richarte, 2015). Moreover, antibody preparation time, poor stability and the effect of modification on antibody binding limit their adaptation in rapid formats. In contrast, nucleic acid aptamers, offer advantages over antibodies for ligand capture including but not limited to: **a)** custom synthesis resulting in practically no batch-to-batch variation; **b)** unlimited shelf life when stored as a dry lyophilized reagent at room temperature (*i.e.*, no requirement for a refrigerator); **c)** easy identification of non-competitive pair of aptamers (aptamer candidates than binds at two distinct sites of same molecules). **d)** Well defined single site conjugation chemistry **e)** Ability to tailor desired affinity and specificity even after SELEX (Post-SELEX modification and affinity maturation) **f)** In cases

where antigens (Ag) are present as a part of immuno-complex (Ag complex with Antibody) an aptamer can be an ideal diagnostic reagent for antigen detection because being a synthetic reagent no such antibody is present against these aptamers and **g)** Due to well established synthesis protocol and ease in synthesis aptamers are cost effective diagnostic reagents and an estimated cost of aptamer is under US\$2,000/g and less than a cent per test (Blind and Blank, 2015; Kedzierski et al., 2013; Ozer et al., 2014). Aptamers have recently celebrated 25th anniversary of their discovery and since inception aptamers have offered a great opportunity to be explored in various areas of science and technology including, but not limited to, diagnostics, imaging, drug delivery, therapeutics, biosensing, environmental monitoring and analytical applications (Gopinath et al., 2016). Moreover, due to the ability to bind their targets tightly and specifically various aptamers have been utilized in immunoassays and other related diagnostic formats (Gopinath et al., 2016).

The advent of point-of-care diagnostics (POCD) has spurred a great interest among researchers, clinicians and health care providers due to the increased speed of diagnostics without compromising the diagnostic quality and accuracy that can be provided at a lower price (Pai et al., 2012; Xiang and Lu, 2012). Innovation in rapid POCD tests creates several next generation assays that can analyze blood sugar levels, metabolites and can determine pregnancy within a few minutes using appropriate body fluids (Gopinath et al., 2016; Pai et al., 2012; Xiang and Lu, 2012). Similarly, on-site contamination of water with microbes or metal ions can easily be detected. This allows the rapid communication of results followed by necessary therapeutic interventions on the very first clinical encounter. Pai et al. (2012) suggested that the speed of a POC test followed by communication of results to the clinicians are the two most critical parameters that govern the success of a POC test. Though an ideal POC test may follow the “ASSURED” criteria: affordable, sensitive, specific, user friendly, robust, and deliverable and perform outside the laboratory but even if a diagnostic test is not following one or more of such criteria but delivering the results and communicating the information in a single day, then also that particular test must be recognized as a POC test as it is fulfilling the diagnostic need (Pai et al., 2012). Therefore, any test homogenous (mix and read, no immobilization and washing are required) or heteroge-

neous (immobilization and washing are required) that can be completed within a day and disseminate the result to the clinician on same day can be regarded as a POC test and may include lateral flow, Fluorescence Resonance Energy Transfer (FRET) and non-FRET assays, Enzyme-Linked Immunosorbent Assay (ELISA) or Aptamer-Linked Immunosorbent Assay (ALISA), Apt-PCR (aptamer-mediated polymerase chain reaction), aptamer-mediated electrochemical and nanoparticles based assays. Fortunately, aptamers work well in all of the aforementioned and related formats and have the ability to compete against antibodies very strongly with the potential to replace antibodies in all possible point-of-care formats and it is evident by a large number of published reports where detection of diseases associated protein biomarkers and small molecules are demonstrated using aptamers but unfortunately a clear translation from research to product is somehow missing (Bruno, 2009, 2012c, 2014, 2015; Chen and Yang, 2015). We herein discuss potential strategies that may enhance “bench to bedside” or “research to product” translation.

Potential of Aptamers and Aptamer Industry

The market potential of aptamer is continuously growing in various applications including but not limited to diagnostics, bio-sensing, biomarker discovery, drug discovery therapeutics and research tool development. We also believe that the diagnostic ability of nucleic acid aptamers is not yet explored to its fullest. The possible reason could be the huge investment and thus commitment of diagnostic companies for the production of antibodies (poly or mono-clonal) followed by their reluctant and ignorant behaviour towards aptamers and the advantages they offer (Baird, 2010). Moreover, even research laboratories hesitate to replace antibodies with aptamers despite knowing their advantages probably due to fear of trying new technology and replacement of their optimized protocols with newer one or may be due to anxiety of spending time on optimization (Baird, 2010). If we compare the translation aspect of aptamers with that of monoclonal antibodies (mAb), it becomes clear that mAbs also took almost 4 decades to reach real translation (Bruno, 2015). Though in terms of translation aptamers seem to be lagging behind antibodies. However, some serious efforts made by aptamer companies cannot be ignored. Approximately 50 aptamer companies are actively engaged in aptamer research with prime focus on at least

one of the following areas: diagnostics, therapeutics,

Table 1: List of aptamer companies and their country of location

S.N.	Company	Country of location
1	AptaBharat Innovation	India
2	Aptahem	Sweden
3	AptaBiosciences	Singapore & UK
4	Apta Biotherapeutics	South Korea
5	Aptamer Sciences	South Korea
6	Apta Targets	Spain
7	AptusBiotech	Spain
8	AptaIT	Germany
9	AuramerBio	Newzeland
10	Apterna	UK
11	Aptamer Group	UK
12	amsBio	UK
13	ATDBio	UK
14	Aptadx	UK
15	Aptamer Solutions	UK
16	AstraZeneca	UK
17	AMBiotech	USA
18	Aptitude Medical	USA
19	Aptagen	USA
20	Aptamatrix	USA
21	Alpha Diagnostic International	USA
22	Avacta	USA
23	Archemix	USA
24	Altermune	USA
25	BBi Group	UK
26	BasePair Technologies	USA
27	CD genomics	USA
28	Firefly Bioworks	USA
29	Ice9Biotechnologies	USA
30	Izon Science	UK
31	Iba Gmbh	Germany
32	LFB Biotech	France
33	LC Science	USA
34	Novaptech	France
35	Neovventure Biotech.	Canada
36	Nal von Minden	Germany
37	Ophthotech	USA
38	OTC Biotech	USA
39	Oak Biosciences	USA
40	NOXXON Pharma Ag	Germany
41	Piculet Biosciences	Netherland
42	Pure biologics	Poland
43	Ribomic	Japan

44	Somallogic	USA
45	Tobira Therapeutics	USA
46	Tocris Bioscience	USA
47	Trilink Biotech	USA
48	Vivonics	USA
49	Vivonics	USA
50	Veraptus	China

and development of novel technologies for various applications. Though developed countries are leading the field in terms of aptamer industry, developing countries including Asian subcontinent (India, China and South Korea) are also swiftly contributing to the growth of aptamer industry. A comprehensive list of various aptamer companies and their location is tabulated in Table 1. Among these aptamer companies many of them have been established in the last 5-6 years, some major players include, but are not limited to, NeoVentures Biotechnologies (Canada), OTC Biotech (USA), Base Pair Biotechnologies (USA), Aptagen (USA), Aptamer Sciences (Pohang, Korea), the Aptamer Group (U.K.) and a few others. Neo Ventures showed great promise by developing and commercialization of their OTA-sense and Afla-sense technology, an aptamer based concentration and purification column for the detection of ochratoxin and aflatoxin respectively in corn, wheat, beer and wines (Penner, 2012). Aptagen developed Apta-Beacon technology for bio-sensing. Aptamer Sciences has a range of aptamer products under their trademark including AptoCyto and a variety of aptamers against plethora of targets for flow cytometry application. The remainders of the aptamer companies are primarily focusing on development of the aptamer-based research tools such as development of aptamers as a substitute of antibodies for the detection of small molecules, proteins, transcription factors, bacterial and viral pathogens protein purification and western blot applications. The Aptamer Group recently launched their technology called Aptabind, an aptamer-based therapeutics and protein purification technology based on aptamer-mediated chromatography. Another major aptamer player called SomaLogic (USA), has developed the SOMAScan platform, an aptamer array that has capability to detect more than 1,000 proteins down to subpicogram levels in serum and other body fluids and has already identified biomarkers for various diseases such as tuberculosis, prostate cancer and more recently for idiopathic pulmonary fibrosis (Kraemer et al., 2011; Nahid et al., 2014; Rohloff et al., 2014).

This technology has paved the new way in the diagnostic arena and can be utilized to identify both novel pathogen or host associated markers for diagnostic and treatment monitoring. These aforementioned achievements are quite encouraging for the field but many more such translations are yet to come in order to swiftly lead to aptamer commercial success.

Strategies to Improve the Translation of Aptamers

In order to achieve the successful “bench to bedside” translation an aptamer must follow the 4 rules as described by Penner (2012): (1) The developed aptamer-based product/assay must be cheaper than the existing product. However, if no such competitive test/assay is available in the market then also the developed test must be cost effective so that the benefit to the user can be maximized. (2) The developed test must exhibit same level of sensitivity regardless of batch. (3) The test should show a high level of sensitivity so as to match the country-specific regulatory standards and (4) the developed test must clearly demonstrate advantages (one or more) over the existing immunoassay products. In our opinion one more rule can be added to this list as a 5th rule: If the aptamer-based assay is integrated with an instrument then this instrument must be more portable and affordable. Apart from these rules for a successful “bench to bedside” translation of aptamer research one should clearly define the target product profile (TPP) or end user that are going to use the test at the very beginning of research which will certainly minimize labor and post-development modifications and alterations. There are five TPP that are helpful at various levels like TPP1 (home/industry), TPP2 (communities),

TPP3 (clinics), TPP4 (diagnostics labs or labs present in industry settings) and TPP5 (hospitals) (Denkinger and Pai, 2012; Pai et al., 2012). Pre-defining the TPP may increase the speed of success and can help in designing a diagnostic prototype matching the end user requirements. However, as such no one has defined the TPP for aptamer research.

We herein made an attempt to define TPPs for aptamer research. Since aptamers are equally applicable for clinical diagnostics and food industry setting (quality control), Thus, we have included both diagnostics and industry application (Figure 1). Further, it is advisable that instead of creating a new prototype, which is quite challenging and time-consuming. If you can combine your aptamer/aptasensors with an existing routinely used well-accepted device, it will certainly enhance the speed of translation and may enable omission of various steps including the design manufacturing and validation of the prototype. One such example is the pioneering work of Lu (2011) where he and his colleague combine the sensing ability of aptamers with a routinely used Personal Glucose Monitor (PGM) device by conjugating the analyte specific aptamer with invertase, an enzyme that converts sucrose to glucose and fructose. The authors accurately detected interferon-gamma, cocaine, uranium and adenosine down to the micro (cocaine and adenosine) and nanomolar ranges (Lu, 2011). In another study, a colorimetric cocaine aptasensor was coupled to an android application resulting in smart phone-based bio-sensing that is again a good example of aptamer-enabled applications of daily use “gadgets” for bio-sensing (Smith et al., 2014).

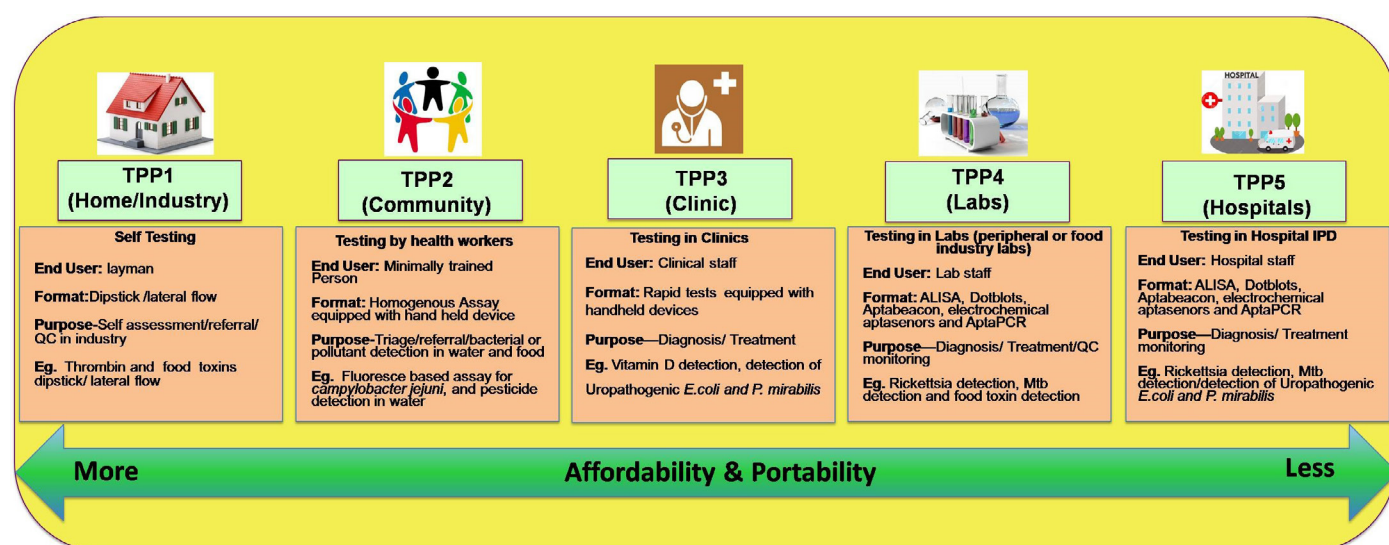


Figure 1: Representation of Target product Profiles (TPPs) for aptamer research. The concept of TTPs is adapted from Pai et al. (2012)

The other strategy for successful translation of aptasensors includes the development of “bind and detect” homogenous assays to address the specific need where antibody cannot perform or poorly performs (Bruno, 2009, 2012b, 2015). Such assays can clearly demonstrate the advantage of aptamers vs. antibodies and may divert the attention of the research community towards aptamers. This FRET assay is reported by Bruno et al. (2011a), (2011b) and (2012) for the precise quantification of microgravity-induced urinary bone loss markers and serum vitamin D levels (Bruno et al., 2001, 2008, 2010, 2011, 2012; Ilgu et al., 2016). This FRET assay can be coupled to commercially available QuantiFluor™ (Promega Corp.) or any other portable fluorometer to address the diagnostic need in resource-limited settings (Bruno et al., 2011, 2012). Similarly, Bruno et al. (2009) developed a novel plastic-adherent aptamer-based quantum dot-based fluorescent assay for the detection of *Campylobacter jejuni* (Bruno et al., 2009). Furthermore, the anti-stokes ability of lanthanide upconverting nanophosphors / nanoparticles (UCNPs) may be exploited in combination with aptamers to develop an affordable “bind and detect” homogenous assay for clinical application. The anti-stokes ability of these UCNPs can reduce the autofluorescence (background fluorescence) problem commonly encountered when dealing with biological fluids (Duan et al., 2012; Fang et al., 2014; Wu et al., 2014). For successful translation of technology, both researchers and entrepreneurs must focus on the development of more handy platforms like lateral flow/dip stick formats. This will certainly enhance the image of aptamers in health care sectors and make them more popular for TPP1. Researchers have already achieved a few leads in this direction, like aptamer-based lateral flow test strips for thrombin, aflatoxin, ochratoxin, cocaine, adenosine and *E.coli* (Chen and Yang, 2015). It is high time to be capitalizing on these leads to make a successful “bench to bed side” translation. In order to match the sensitivity of regulatory standards multivalent aptamer constructs may be a good idea since in several studies they have shown promising improvements in aptamer affinity and sensitivity. Also, we envision that one should test the developed aptamers in all possible formats (lateral flow, beacons, electrochemical, ALISA etc.) so as to get an appropriate platform to work for a diagnostic test.

Future of Aptamer-based Diagnostics

Aptamers have the ability to perform in almost every diagnostic format including homogenous multiplex

assays where antibodies may fail or poorly perform suggesting that in the future there is a fair chance to replace antibodies with aptamers in diagnostics. A PubMed search for “aptamer in diagnostic” on August 29, 2016, yielded 1477 hits, while phrase “aptamer in bio-sensing” resulting 1456 hits. These large numbers of hits seem to indicate that the field of aptamers is flourishing in the arenas of diagnostics and bio-sensing. This also suggests opportunity for capitalizing on fruitful aptamer-based diagnostic products that work exceptionally well. Moreover, based on the active engagement of several companies in aptamer development with a main focus on diagnostics and bio-sensing streams, we envision a bright future for aptamers in the diagnostic market. The ease of scale up and speed of aptamer development would certainly contribute to success of aptamer-based diagnostics. One cautionary note from our side is that one should also evaluate the stability of aptamers in diagnostic format, which would certainly build confidence in the developer and also in the end user. Also, the aptamer community should choose more clinically relevant aptamer targets so as to convert aptamer leads into acceptable and much needed products. This will certainly overcome the hesitation of lab users and clinicians generated due to entrenched antibody-based assays. Then aptamers should come up as next generation reagents in the molecular tool kit of diagnostic labs.

Acknowledgments

T.K.S. wish to thank Department of Biotechnology, Govt. of India for Innovation Award.

Conflict of Interest

T.K.S. owns 65% stake in AptaBharat Innovation Pvt. Ltd. India and JGB owns 8% stakes in OTC Biotechnology USA.

Authors' Contribution

All authors have contributed in writing and revising the manuscript.

References

- Baird, G.S. (2010). Where Are All the Aptamers? *Am J Clin Pathol*, 134, 529-531. <http://dx.doi.org/10.1309/AJCPFU4CG2WGJJKS>
- Blind, M., and Blank, M. (2015). Aptamer Selec-

- tion Technology and Recent Advances. *Mol Therap Nucl Acids*, 4(1), e223. <http://dx.doi.org/10.1038/mtna.2014.74>
- Bruno, J.G. (2014). Application of DNA Aptamers and Quantum Dots to Lateral Flow Test Strips for Detection of Foodborne Pathogens with Improved Sensitivity versus Colloidal Gold. *Pathogens*, 3(2), 341-355. <http://dx.doi.org/10.3390/pathogens3020341>
 - Bruno, J.G. (2015). Predicting the Uncertain Future of Aptamer-Based Diagnostics and Therapeutics. *Molecules*, 16(20), 6866-6887. <http://dx.doi.org/10.3390/molecules20046866>
 - Bruno, J.G., Richarte, A.M., Carrillo M.P., Edge, A. 2012b. An aptamer beacon responsive to botulinum toxins. *Biosens Bioelectron*, 31, 240-243. <http://dx.doi.org/10.1016/j.bios.2011.10.024>
 - Bruno, J.G., Carrillo, M.P. (2012c). Development of aptamer beacons for rapid presumptive detection of Bacillus spores. *J Fluorescence*, 22, 915-924. <http://dx.doi.org/10.1007/s10895-011-1030-0>
 - Bruno, J.G., Phillips, T., Carrillo, M.P., Crowell, R. (2009). Plastic-adherent DNA aptamer-magnetic bead and quantum dot sandwich assay for *Campylobacter* detection. *J Fluorescence*, 19, 427-435. <http://dx.doi.org/10.1007/s10895-008-0429-8>
 - Bruno, J.G., Carrillo, M.P., Phillips, T., Andrews, C.J. (2010). A novel screening method for competitive FRET-aptamers applied to *E. coli* assay development. *J Fluoresc*, 20(6), 1211-1223. <http://dx.doi.org/10.1007/s10895-010-0670-9>
 - Bruno, J.G., Carrillo, M.P., Phillips, T., Edge, A. (2012). Serum inverts and improves the fluorescence response of an aptamer beacon to various vitamin D analytes. *Luminescence*, 27(1), 51-58. <http://dx.doi.org/10.1002/bio.1324>
 - Bruno, J.G., Carrillo, M.P., Phillips, T., Vail, N.K., Hanson, D. (2008). Competitive FRET-aptamer-based detection of methylphosphonic acid, a common nerve agent metabolite. *J Fluoresc*, 18(5), 867-876. <http://dx.doi.org/10.1007/s10895-008-0316-3>
 - Bruno, J.G., Phillips, T., Carrillo, M.P., Crowell, R. (2009). Plastic-adherent DNA aptamer-magnetic bead and quantum dot sandwich assay for *Campylobacter* detection. *J Fluoresc*, 19(3), 427-435. <http://dx.doi.org/10.1007/s10895-008-0429-8>
 - Bruno, J.G., Richarte, A. (2015). Aptamer-Quantum Dot Lateral Flow Test Strip Development for Rapid and Sensitive Detection of Pathogenic *Escherichia coli* via Intimin, O157- Specific LPS and Shiga Toxin 1 Aptamers. *Curr Bionanotechnol*, 1, 80-86. <http://dx.doi.org/10.2174/2213529402999160502115500>
 - Bruno, J.G., Ulvick, S.J., Uzzell, G.L., Tabb, J.S., Valdes, E.R., Batt, C.A. (2001). Novel immuno-FRET assay method for *Bacillus* spores and *Escherichia coli* O157:H7. *Biochemical Biophysical Res Commun*, 287(4), 875-880. <http://dx.doi.org/10.1006/bbrc.2001.5669>
 - Bruno, J.G., Zuniga, M.A., Carrillo, M.P., Phillips, T. (2011). Development of naturally selected and molecularly engineered intrachain and competitive FRET-aptamers and aptamer beacons. *Comb Chem High Throughput Screen*, 14(7), 622-630. <http://dx.doi.org/10.2174/138620711796367175>
 - Chen, A., Yang, S. (2015). Replacing antibodies with aptamers in lateral flow immunoassay. *Biosens Bioelectron*, 71, 230-242. <http://dx.doi.org/10.1016/j.bios.2015.04.041>
 - Chopra, A., Shukla, R., Sharma, T.K. (2014). Aptamers as an Emerging Player in Biology. *Aptamer Synthet Antibodies*, 1, 1-11.
 - Citartan, M., Gopinath, S.C., Tominaga, J., Tan, S.C., Tang, T.H. (2012). Assays for aptamer-based platforms. *Biosens Bioelectron*, 34(1), 1-11. <http://dx.doi.org/10.1016/j.bios.2012.01.002>
 - Denkinger, C.M., Pai, M. (2012). Point-of-care tuberculosis diagnosis: are we there yet? *The Lancet. Infect Dis*, 12(3), 169-170. [http://dx.doi.org/10.1016/s1473-3099\(11\)70257-2](http://dx.doi.org/10.1016/s1473-3099(11)70257-2)
 - Duan, N., Wu, S., Zhu, C., Ma, X., Wang, Z., Yu, Y., Jiang, Y. (2012). Dual-color upconversion fluorescence and aptamer-functionalized magnetic nanoparticles-based bioassay for the simultaneous detection of *Salmonella Typhimurium* and *Staphylococcus aureus*. *Analytica Chimica Acta*, 723, 1-6. <http://dx.doi.org/10.1016/j.aca.2012.02.011>
 - Fang, S., Wang, C., Xiang, J., Cheng, L., Song, X., Xu, L., Peng, R., Liu, Z. (2014). Aptamer-conjugated upconversion nanoprobe assisted by magnetic separation for effective isolation and sensitive detection of circulating tumor cells. *Nano Res*, 7(9), 1327-1336. <http://dx.doi.org/10.1007/s12274-014-0497-9>
 - Gopinath, S.C., Lakshmi Priya, T., Chen, Y., Phang, W.M., Hashim, U. (2016). Aptamer-based 'point-of-care testing'. *Biotechnol Adv*, 34(3), 198-208. <http://dx.doi.org/10.1016/j.biotechadv.2016.02.003>
 - Ilgu, M., Ray, J., Bendickson, L., Wang, T., Geraskin, I.M., Kraus, G.A., Nilsen-Hamilton, M.

- (2016). Light-up and FRET aptamer reporters; evaluating their applications for imaging transcription in eukaryotic cells. *Methods*, 98, 26-33. <http://dx.doi.org/10.1016/j.ymeth.2015.12.009>
- Kedzierski, S., Khoshnejad, M., Caltagirone, G.T. (2013). Synthetic antibodies: The emerging field of aptamers. *BioProcess J Winter*, 2012-2013, 46-49. <http://dx.doi.org/10.12665/j114.kedzierski-caltagirone>
 - Kraemer, S., Vaught, J.D., Bock, C., Gold, L., Katilius, E., Keeney, T.R., Kim, N., Saccomano, N.A., Wilcox, S.K., Zichi, D., Sanders, G.M. (2011). From SOMAmer-based biomarker discovery to diagnostic and clinical applications: a SOMAmer-based, streamlined multiplex proteomic assay. *PLoS One*, 6(10), e26332. <http://dx.doi.org/10.1371/journal.pone.0026332>
 - Lange, M.J., Sharma, T.K., Whatley, A.S., Landon, L.A., Tempesta, M.A., Johnson, M.C., Burke, D.H. (2012). Robust suppression of HIV replication by intracellularly expressed reverse transcriptase aptamers is independent of ribozyme processing. *Mol Ther*, 20(12): 2304-2314. <http://dx.doi.org/10.1038/mt.2012.158>
 - Xiang, Y., Lu, Y. (2011). Using personal glucose meters and functional DNA sensors to quantify a variety of analytical targets. *Nature Chem*, 3: 697-703.
 - Nahid, P., Bliven-Sizemore, E., Jarlsberg, L.G., De Groote, M.A., Johnson, J.L., Muzanyi, G., Engle, M., Weiner, M., Janjic, N., Sterling, D.G., Och-sner, U.A. (2014). Aptamer-based proteomic signature of intensive phase treatment response in pulmonary tuberculosis. *Tuberculosis*, 94(3), 187-196. <http://dx.doi.org/10.1016/j.tube.2014.01.006>
 - Ozer, A., Pagano, J.M., Lis, J.T. (2014). New Technologies Provide Quantum Changes in the Scale, Speed, and Success of SELEX Methods and Aptamer Characterization. *Mol Ther Nucl Acids*, 3, e183. <http://dx.doi.org/10.1038/mtna.2014.34>
 - Pai, N.P., Vadnais, C., Denking, C., Engel, N., Pai, M. (2012). Point-of-care testing for infectious diseases: diversity, complexity, and barriers in low- and middle-income countries. *PLoS Med*, 9(9), e1001306. <http://dx.doi.org/10.1371/journal.pmed.1001306>
 - Rohloff, J.C., Gelinas, A.D., Jarvis, T.C., Och-sner, U.A., Schneider, D.J., Gold, L., Janjic, N. (2014). Nucleic acid ligands with protein-like side chains: Modified aptamers and their use as diagnostic and therapeutic agents. *Mol Ther Nucl Acids*, 3, e201. <http://dx.doi.org/10.1038/mtna.2014.49>
 - Sharma, T.K., Ramanathan, R., Weerathunge, P., Mohammadtaheri, M., Daima, H.K., Shukla, R., Bansal, V. (2014). Aptamer-mediated 'turn-off/turn-on' nanozyme activity of gold nanoparticles for kanamycin detection. *Chem Commun*, 50(100), 15856-15859. <http://dx.doi.org/10.1039/C4CC07275H>
 - Sharma, T.K., Shukla, R. (2014a). Nanoscience and aptamer technology for point of care diagnostics. *Nanosci Technol*, 1, 1-2.
 - Sharma, T.K., Shukla, R. (2014b). Nucleic acid aptamers as an emerging diagnostic tool for animal Pathogens. *Adv Anim Vet Sci*, 2, 50 – 55. <http://dx.doi.org/10.14737/journal.aavs/2014.2.1.50.55>
 - Smith, J.E., Griffin, D.K., Leny, J.K., Hagen, J.A., Chavez, J.L., Kelley-Loughnane, N. (2014). Col-orimetric detection with aptamer-gold nanoparticle conjugates coupled to an android-based color analysis application for use in the field. *Talanta*, 121, 247-255. <http://dx.doi.org/10.1016/j.talanta.2013.12.062>
 - Kedzierski, S., Khoshnejad, M., Caltagirone, G.T., 2013. Synthetic antibodies: The emerging field of aptamers. *Bioprocess J*, 11: 46-49.
 - Tuerk, C., Gold, L. (1990). Systematic Evolution of Ligands for Exponential Enrichment: RNA Ligand to Bacteriophage T4 DNA Polymerase. *Science*, 249: 505-10 <http://dx.doi.org/10.1126/science.2200121>
 - Weerathunge, P., Ramanathan, R., Shukla, R., Sharma, T.K., Bansal, V. (2014). Aptamer-controlled reversible inhibition of gold nanozyme activity for pesticide sensing. *Analytical Chem*, 86(24), 11937-11941. <http://dx.doi.org/10.1021/ac5028726>
 - Wu, S., Duan, N., Shi, Z., Fang, C., Wang, Z. (2014). Simultaneous aptasensor for multiplex pathogenic bacteria detection based on multi-color upconversion nanoparticles labels. *Analytical Chem*, 86(6), 3100-3107. <http://dx.doi.org/10.1021/ac404205c>
 - Xiang, Y., Lu, Y. (2012). Portable and quantitative detection of protein biomarkers and small molecular toxins using antibodies and ubiquitous personal glucose meters. *Analytical Chem*, 84(9), 4174-4178. <http://dx.doi.org/10.1021/ac300517n>