



# Systematic and Integrated Analysis Approach to Prioritize Mastitis Resistant Genes

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## ABSTRACT

Mastitis (mammary gland inflammation) is one of the most important diseases causing economic losses to dairy farm owners. In order to develop new strategies to prevent mastitis, a detailed understanding of the molecular mechanisms underlying the host immune response to the infectious agents is necessary. Identification of the genes, and their variants, involved in innate immune responses is essential for the understanding of this inflammatory disease and to identify potential genetic markers for resistance to mastitis. This article presents a systematic integration of complex biological interactions of 226 mammary gland genes to uncover underlying regulatory networks. Data were collected from various databases after having an exhaustive literature study. Network, functional and pathway analysis of the mammary gland genes were performed to find mastitis resistant genes, which helped us to identify prime genes of mammary gland that are more resistant to mastitis, and analysis of those resistant genes helped to find the solution of mastitis and immunity related diseases.

## Article Information

Received 03 August 2016

Revised 28 August 2016

Accepted 07 September

Available online 02 November 2016

## Authors' Contributions

AN and MMA conceived the study; IM, AN, MEB, MMA, MJ, AS, TH and MTP designed and performed the experiments. AN, MJ, IM drafted the article.

## Key words

Systematics, Inflammatory diseases, Intra-mammary infection, Mammary glands, Immunity, Bovine genome, Networks, Pathways, Mastitis resistance genes

## INTRODUCTION

Worldwide human consumption of bovine milk and by-products has increased in the last decades, especially in countries where this food was not part of the traditional diet (Wiley, 2007). The bovine mammary gland is an extraordinary organ able to produce 30,000 kg of milk in a complete lactation cycle (Bionaz *et al.*, 2012). Major advances have been seen in the past several decades in understanding the physiology of lactating mammary gland driven partly by the desire to improve the efficiency of production and partly by the desire to increase quality of the milk (Bauman *et al.*, 2006). The development of high throughput tools such as microarray analysis has provided the unique feature to uncover the molecular networks in mammary tissue during the course of pregnancy, milk synthesis, involution and diseases related to mammary gland *e.g.*, mastitis (Rudolph *et al.*, 2003). However, interpretation of this large amount of data was extremely challenging. The advent of bioinformatics tools has allowed summarizing large microarray data sets, revealing an integral view of cellular phenomena (Lemay *et al.*, 2007). Mastitis is an inflammatory disease of the bovine mammary gland that occurs in response to physical damage or infection with pathogenic microorganisms. Infection of the mammary gland causes macrophages and epithelial cells to release cytokines which cause polymorphonuclear neutrophils (PMNs), monocytes, and other leukocytes to migrate from the blood to the site of infection in the mammary tissue (Schukken *et al.*, 2011; Paape *et al.*, 2003). This influx of

leukocytes results in an increased level of somatic cells in the milk. It costs more than \$2 billion alone in USA (Viguier *et al.*, 2009). In Pakistan only in Punjab Rs. 240 million is the average loss due to mastitis per year (Khan and Khan, 2006). It usually occurs primarily in response to intra-mammary bacterial infection but mycoplasmal, fungal, or algal infections are also causes of mastitis among which *Staphylococcus aureus* and *Escherichia coli* are the most important gram-positive and gram-negative bacteria, respectively (Schukken *et al.*, 2011). The prevalence of mastitis in dairy cattle is relatively high.

Keeping in view the importance of milk products and loss due to mastitis, there is a need to develop new and improved control strategies. For this purpose a detailed understanding of the molecular mechanisms underlying the host immune response to the infectious agents is necessary (Jensen *et al.*, 2013). Although the differential expression of genes studied through microarray technique has helped to understand the gene networks and functional pathways affected most in mammary tissue in response to an intra-mammary infection with *S. uberis* (Moyes *et al.*, 2009), identification of the genes and their variants involved in innate immune responses is essential for the understanding of this inflammatory disease and to identify potential genetic markers for resistance to mastitis in bovine. This article presents analytical study of bovine mammary gland genes to uncover underlying regulatory networks.

## MATERIALS AND METHODS

### Literature survey and data collection

This article presents analytical study of bovine mammary gland genes to uncover underlying regulatory networks. Network, functional and pathway analysis of 226 mammary

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0030-9923/2016/0004-1161 \$ 8.00/0

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gland genes were performed to find mastitis resistant genes. Data was collected from UniGene, NCBI, Ensembl and bovine genome databases. Network analysis was performed by STRING (Search Tool for the Retrieval of Interacting Genes/Proteins). For functional analysis DAVID (the Database for Annotation, Visualization and Integrated Discovery) was used and pathway analysis was performed through KEGG (Kyoto Encyclopedia of Genes and Genomes) (Kanehisa, 2013).

A list of genes representing part of the mammalian mammary gland transcriptome was assembled by an extensive literature survey and searching various databases. The UniGene which is an NCBI database of the transcriptome was searched to identify genes expressed in bovine mammary gland. Literature published was reviewed by searching for the relevant publications through PubMed using key phrases: association, gene candidates, epigenetics, genetics, mammary gland, mastitis, methylation, milk, miRNA, QTL, SNP. A systematic review of both original research articles and reviews was performed for searching literature associated with bovine mammary gland genes. Data of genes was retrieved from the NCBI, Ensembl and bovine genome databases.

#### Network analysis

Complete knowledge of all direct and indirect interactions between proteins in a given cell would represent an important milestone towards a comprehensive

description of cellular mechanisms and functions. Network analysis of the list of mammary gland genes was completed by STRING. It sorted out our data according to predicted functional association among different genes and proteins. We used four views for our analysis which were, Co-expression, database, experiment and text mining at high confidence score of 0.700. This analysis helped in systematically making the group of genes performing similar functions and showing similar expression pattern.

#### Functional analysis

DAVID was used to perform analysis of functionally related genes. Functional annotation and gene functional classification tools were used for the functional analysis of mammary gland genes at  $p < 0.05$ . This gave us an in-depth understanding of the biological themes in list of genes that are enriched in genome-scale studies.

#### Pathway analysis

Pathway analysis of mastitis resistant genes was performed through KEGG which gave an insight into pathways where the genes were involved and it helped to interpret the data in the context of biological processes, pathways and networks. This analysis helped to understand the mechanisms of mastitis, the unaffected pathways and the pathways that were affected most. These findings ultimately helped to find resistance mechanism and genes involved in resistance.

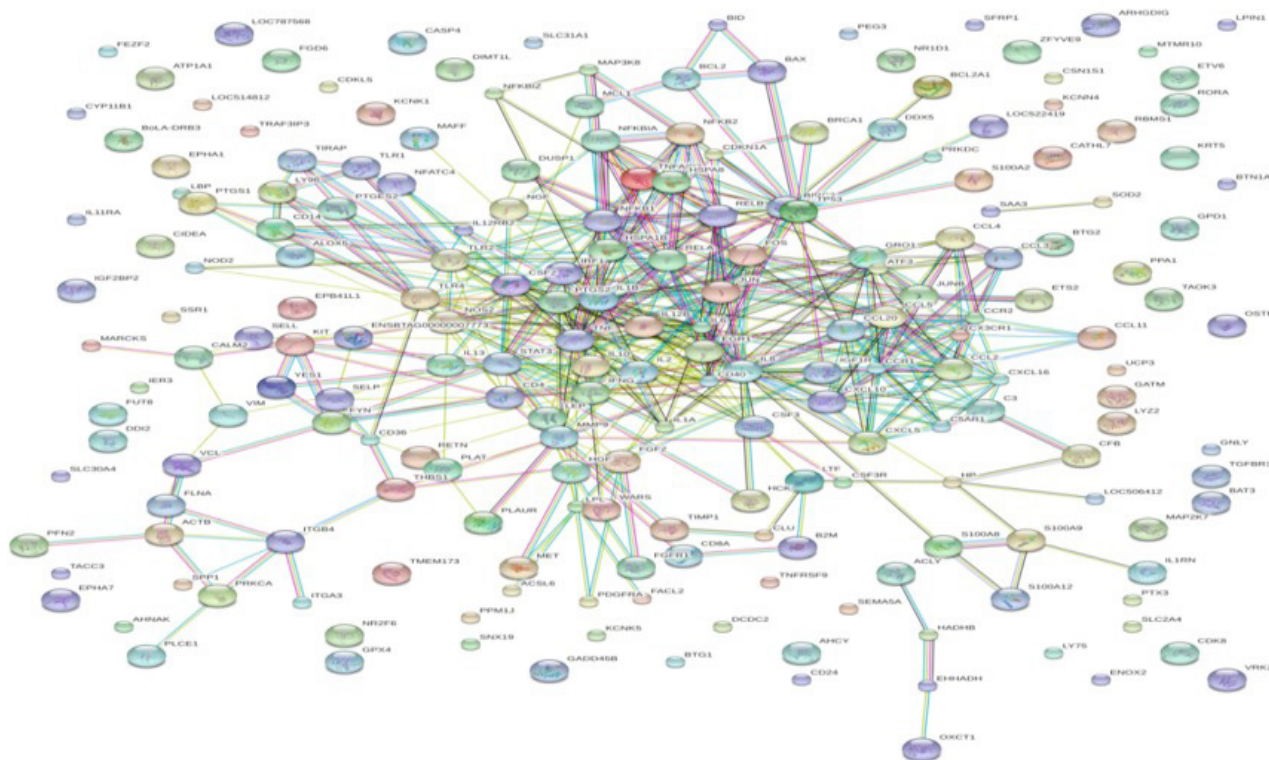


Fig. 1. The network showing 3 clusters: In the center there is the largest network of the dataset; in the middle there is a second largest network of genes and at the sides scattered genes are shown that are not interconnected to any other gene in the provided dataset.

**Table I.- Gene functional classification.**

| <b>Gene Group 1</b>  | <b>Enrichment Score: 14.712468824680236</b>                                                                                                                                                            |
|----------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| CCL11                | chemokine (C-C motif) ligand 11                                                                                                                                                                        |
| IL13                 | interleukin 13                                                                                                                                                                                         |
| IL8                  | interleukin 8                                                                                                                                                                                          |
| CCL2                 | chemokine (C-C motif) ligand 2                                                                                                                                                                         |
| CCL3                 | chemokine (C-C motif) ligand 3                                                                                                                                                                         |
| CCL5                 | chemokine (C-C motif) ligand 5                                                                                                                                                                         |
| CXCL16               | chemokine (C-X-C motif) ligand 16                                                                                                                                                                      |
| CXCL5                | chemokine (C-X-C motif) ligand 5                                                                                                                                                                       |
| CSF3                 | colony stimulating factor 3 (granulocyte)                                                                                                                                                              |
| CCL4                 | chemokine (C-C motif) ligand 4                                                                                                                                                                         |
| IL1RN                | interleukin 1 receptor antagonist                                                                                                                                                                      |
| CCL20                | chemokine (C-C motif) ligand 20                                                                                                                                                                        |
| CXCL10               | chemokine (C-X-C motif) ligand 10                                                                                                                                                                      |
| <b>Gene Group 2</b>  | <b>Enrichment Score: 6.197470798536051</b>                                                                                                                                                             |
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| IFNG                 | interferon, gamma                                                                                                                                                                                      |
| IL10                 | interleukin 10                                                                                                                                                                                         |
| IL1B                 | interleukin 1, beta                                                                                                                                                                                    |
| IL2                  | interleukin 2                                                                                                                                                                                          |
| TNF                  | tumor necrosis factor (TNF superfamily, member 2)                                                                                                                                                      |
| <b>Gene Group 3</b>  | <b>Enrichment Score: 3.3291366663923707</b>                                                                                                                                                            |
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| RELA                 | v-relreticuloendotheliosis viral oncogene homolog A (avian); similar to v-relreticuloendotheliosis viral oncogene homolog A, nuclear factor of kappa light polypeptide gene enhancer in B-cells 3, p65 |
| NFKB2                | nuclear factor of kappa light polypeptide gene enhancer in B-cells 2 (p49/p100)                                                                                                                        |
| NFKB1                | nuclear factor of kappa light polypeptide gene enhancer in B-cells 1                                                                                                                                   |
| RELB                 | v-relreticuloendotheliosis viral oncogene homolog B                                                                                                                                                    |
| <b>Gene Group 4</b>  | <b>Enrichment Score: 2.1814765223414283</b>                                                                                                                                                            |
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| IGF1R                | insulin-like growth factor 1 receptor                                                                                                                                                                  |
| YES1                 | v-yes-1 Yamaguchi sarcoma viral oncogene homolog 1                                                                                                                                                     |
| FGFR1                | fibroblast growth factor receptor 1                                                                                                                                                                    |
| MAP3K8               | mitogen-activated protein kinase kinasekinase 8                                                                                                                                                        |
| MET                  | met proto-oncogene (hepatocyte growth factor receptor)                                                                                                                                                 |
| HCK                  | hemopoietic cell kinase                                                                                                                                                                                |
| KIT                  | v-kit Hardy-Zuckerman 4 feline sarcoma viral oncogene homolog                                                                                                                                          |
| <b>Gene Group 5</b>  | <b>Enrichment Score: 1.8117924213734606</b>                                                                                                                                                            |
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| S100A2               | S100 calcium binding protein A2                                                                                                                                                                        |
| S100A8               | S100 calcium binding protein A8                                                                                                                                                                        |
| S100A9               | S100 calcium binding protein A9                                                                                                                                                                        |
| S100A12              | S100 calcium binding protein A12 (calgranulin C)                                                                                                                                                       |
| <b>Gene Group 6</b>  | <b>Enrichment Score: 1.6666772544237716</b>                                                                                                                                                            |
| OFFICIAL_GENE_SYMBOL | Gene Name                                                                                                                                                                                              |
| CX3CR1               | chemokine (C-X3-C motif) receptor 1                                                                                                                                                                    |
| CSAR1                | complement component 5a receptor 1                                                                                                                                                                     |
| CCR2                 | chemokine (C-C motif) receptor 2                                                                                                                                                                       |
| CCR1                 | chemokine (C-C motif) receptor 1                                                                                                                                                                       |

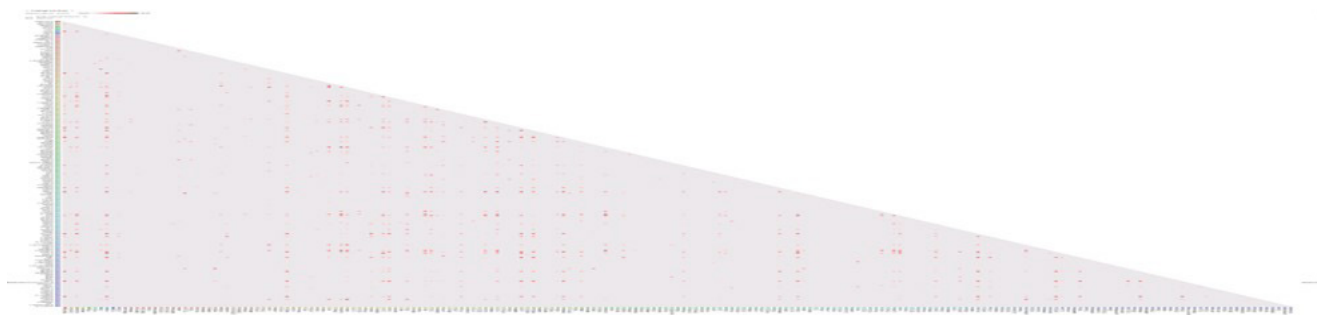


Fig. 2. Co-expressed genes in the dataset. The paler red color represent the less co-expressed genes, the dark red color shows the genes are more co-expressed.

**Table II.- KEGG pathway annotation.**

| Category     | Term                                               |
|--------------|----------------------------------------------------|
| KEGG_PATHWAY | bta04060:Cytokine-cytokine receptor interaction    |
| KEGG_PATHWAY | bta04620:Toll-like receptor signaling pathway      |
| KEGG_PATHWAY | bta04621:NOD-like receptor signaling pathway       |
| KEGG_PATHWAY | bta04660:T cell receptor signaling pathway         |
| KEGG_PATHWAY | bta05200:Pathways in cancer                        |
| KEGG_PATHWAY | bta04010:MAPK signaling pathway                    |
| KEGG_PATHWAY | bta04062:Chemokine signaling pathway               |
| KEGG_PATHWAY | bta04510:Focal adhesion                            |
| KEGG_PATHWAY | bta04640:Hematopoietic cell lineage                |
| KEGG_PATHWAY | bta04210:Apoptosis                                 |
| KEGG_PATHWAY | bta05222:Small cell lung cancer                    |
| KEGG_PATHWAY | bta04920:Adipocytokine signaling pathway           |
| KEGG_PATHWAY | bta05215:Prostate cancer                           |
| KEGG_PATHWAY | bta05218:Melanoma                                  |
| KEGG_PATHWAY | bta04623:Cytosolic DNA-sensing pathway             |
| KEGG_PATHWAY | bta05330:Allograft rejection                       |
| KEGG_PATHWAY | bta04940:Type I diabetes mellitus                  |
| KEGG_PATHWAY | bta04622:RIG-I-like receptor signaling pathway     |
| KEGG_PATHWAY | bta04630:Jak-STAT signaling pathway                |
| KEGG_PATHWAY | bta04520:Adherens junction                         |
| KEGG_PATHWAY | bta05332:Graft-versus-host disease                 |
| KEGG_PATHWAY | bta05020:Prion diseases                            |
| KEGG_PATHWAY | bta05210:Colorectal cancer                         |
| KEGG_PATHWAY | bta05310:Asthma                                    |
| KEGG_PATHWAY | bta04662:B cell receptor signaling pathway         |
| KEGG_PATHWAY | bta04722:Neurotrophin signaling pathway            |
| KEGG_PATHWAY | bta05214:Glioma                                    |
| KEGG_PATHWAY | bta04810:Regulation of actin cytoskeleton          |
| KEGG_PATHWAY | bta05219:Bladder cancer                            |
| KEGG_PATHWAY | bta04610:Complement and coagulation cascades       |
| KEGG_PATHWAY | bta04650:Natural killer cell mediated cytotoxicity |
| KEGG_PATHWAY | bta05220:Chronic myeloid leukemia                  |
| KEGG_PATHWAY | bta04512:ECM-receptor interaction                  |
| KEGG_PATHWAY | bta05221:Acute myeloid leukemia                    |
| KEGG_PATHWAY | bta00590:Arachidonic acid metabolism               |

## RESULTS

### Network analysis

A list of total 226 mammary gland genes was created by systematic and integrated review of literature and provided to STRING. Two hundred and five genes matched with STRING database for *Bos Taurus* and a network was generated of these 205 genes at high confidence score of 0.700 (Fig. 1). One hundred and thirty four genes form the largest network. Figure 2 shows the genes that are co-expressed.

### Functional analysis

DAVID was used for functional analysis of the short listed genes by network analysis. 134 genes that were the part of the largest network were selected for functional analysis to uncover the meaning of interaction of these genes. Functional annotation clustering resulted in a total of 72 clusters. The majority of gene ontology (GO) terms were related to immune response, for example, GO identifiers were related to inflammatory response (GO:0006954), chemotaxis (GO:0006935), immune response (GO:0006955), leukocyte migration (GO:0050900), response to lipopolysaccharide (GO:0032496), and TLR signalling pathway (GO:0002224). To get significant results, we applied an enrichment score cutoff criterion of  $>2$ . The gene list was further sorted out by using another DAVID tool of gene functional classification which sorted the gene list into 6 gene groups as shown in Table I.

### Pathway analysis

Pathway analysis of the functionally related genes was conducted by using KEGG database. Thirty five significant pathways related to these genes were identified among which 11 pathways were directly related to the immunity related pathways. Pathways other than immunity related pathways were apoptosis, small cell lung cancer, adipocytokine signaling pathway, prostate cancer, melanoma, cytosolic DNA-sensing pathway, allograft rejection, type I diabetes mellitus, adherens junction, graft-versus-host disease, prion diseases, colorectal cancer, asthma, glioma, regulation of actin cytoskeleton, bladder cancer, complement and coagulation cascades, chronic myeloid leukemia, ECM-



receptor interaction, acute myeloid leukemia, arachidonic acid metabolism. Total pathways associated with the provided gene list are represented in [Table II](#).

## DISCUSSION

The biological complexity of animals unavoidably requires a systems biology approach, *i.e.*, a way to systematically study the complex biological interactions using a method of integration instead of reduction ([Loor and Cohick, 2009](#)). Important goals of systematic and integrated analysis are to uncover the underlying links (pathways, regulatory networks, and functional organization), and also to discover new emergent properties that may arise from examining the interactions between all components of a system ([Bruggeman and Westerhoff, 2007](#)). In the context of intra-mammary infection (IMI) and the mammary gland response, a “true” systems biology approach would encompass not only the intra-cellular networks of factors, but also communication between tissues ([Piantoni et al., 2010](#)) and between organs/systems.

Most of the genes were related to functions including Immune Response, Immune Disease, Connective Tissue Disorders, Lipid and Carbohydrate Metabolism, Molecular Transport, Cell-To-Cell Signaling, Tissue Development, Cellular Development, and Immune and Lymphatic System Development and Function. These genes encoded cytokines (CSF3, CSF2, CCL3, TNF, CCR1, IL13, KIT, CCL5, CCL4, IL10, CXCL10, IL12RB2, IFNG, IL1B, IL1A, IL8, MET, CD40, HGF, LEP, CXCL16, CX3CR1, CCR2, PDGFRA, IL12B, IL2), lipid metabolism related genes (CD36, GPX4, LPIN1, LPL, LPB), transcription regulators (BCL3, FOS and NFKBIA), receptors (TNF, IL8, RELA, TLR1, TIRAP, TLR2, NFKBIA, NFKB1, TLR4, CD40, CCL5, CXCL10, FOS, JUN, MAP3K8, IL1B, LBP, IL12B, CD14, SPP1), and others such as SELP, SELL, and SOD1, all play a role in some aspect of the immune response including cytokine activity (IL10, TNF, IL8, and IL1B), cell adhesion (SELL and SELP), immune activation (CD14 and TLR2), acute phase reaction (TNF, IL1B, and SAA3), apoptosis (BCL2, BAX). Similar results were observed by Moyes *et al.* (2009) in their study on bovine mammary tissue challenged with *Streptococcus uberis*.

A French group examined the milk somatic cell transcriptome (predominantly PMN) of mastitis susceptible and mastitis-resistant sheep infected successively by *Streptococcus epidermidis* (*S. epidermidis*) and *Staphylococcus aureus* (*S. aureus*) ([Bonnefont et al., 2011](#)). The challenges per se (*i.e.*, *S. aureus* vs. *S. epidermis*) resulted in >5,000 DEG (FDR= 0.05 and fold change >1.5 ratio between *S. aureus* and *S. epidermis*), but the authors performed a functional analysis on those DEG with a >5-fold difference. The functional analysis of genes more expressed in *S. aureus* uncovered significantly-enrichment (EASE score  $\leq 0.05$ ) ([da Huang et al., 2009a](#)), among DEG of functions related to the immune response; whereas, genes more expressed in *S. epidermidis* resulted in over

representation of functions related to metabolism and cell growth, indicating that more T cells were recruited upon inoculation by *S. aureus* than *S. epidermidis*. Few DEG (57 at FDR=0.05 and fold-change >1.5) were found between the mastitis-susceptible and mastitis-resistant sheep. The functional analysis indicated that mastitis-resistant sheep had a more active immune response, as indicated by the significant enrichment of “leukocyte adhesion and activation” among DEG more expressed in mastitis-resistant relative to the mastitis-susceptible sheep ([Bonnefont et al., 2011](#)).

Another German group performed direct transcriptomics comparison between primary bovine mammary epithelial cells isolated from cows with low- or high susceptibility to mastitis challenged either with *E. coli* or *S. aureus* ([Brand et al., 2011](#)). Similarly to the French study mentioned above ([Bonnefont et al., 2011](#)), the main finding from the ORA functional analysis was that the transcriptome of mammary epithelial cells from cows with low-susceptibility to mastitis challenged with either *S. aureus* or *E. coli* had a greater response to the bacterial challenge via up-regulation of genes associated with ‘acute phase response signaling’, *i.e.*, interleukin-6 (IL6) and nuclear factor NF-kappa-B p100 subunit (NFKB2), than cows with high-susceptibility to mastitis ([Brand et al., 2011](#)). Both the German and the French studies appear to support the notion that the sensitivity of the mammary gland to bacteria plays an important role in the resistance to invading microorganisms. Perhaps the evolutionary transition of the mammary gland from a mucus-secreting surface epithelium serving a protective role to a milk-secreting gland with both protective and nutritional roles ([Vorbach et al., 2006](#)) has, as a consequence, a negative impact on the importance of the tissue’s innate immune system.

## CONCLUSION

It was concluded that the stronger immune system of an animal would be more resistant to mastitis. In future, the analysis of this study can be used to further our knowledge about this disease which is causing a huge economical lose worldwide, however, a deeper understanding of its fundamentals and development of more adequate bioinformatics tools is critically needed in order to really benefit from such an approach.

In conclusion, our network-based gene prioritization approach provides a general framework for identifying and ranking genes associated with complex diseases. To our knowledge this is the first time that protein interactions, orthologue mapping, gene expression, and literature mining were integrated for ranking candidate genes in any livestock species. The approaches and techniques that were systematically implemented in the present study are general and not confined by specific trait or species and can be applied to various complex diseases in different organisms.

### Conflict of interest statement

We declare that we have no conflict of interest.

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